

# An antibody based approach for multi-coloring osteogenic and chondrogenic proteins in tissue engineered constructs

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## PAPER

## An antibody based approach for multi-coloring osteogenic and chondrogenic proteins in tissue engineered constructs

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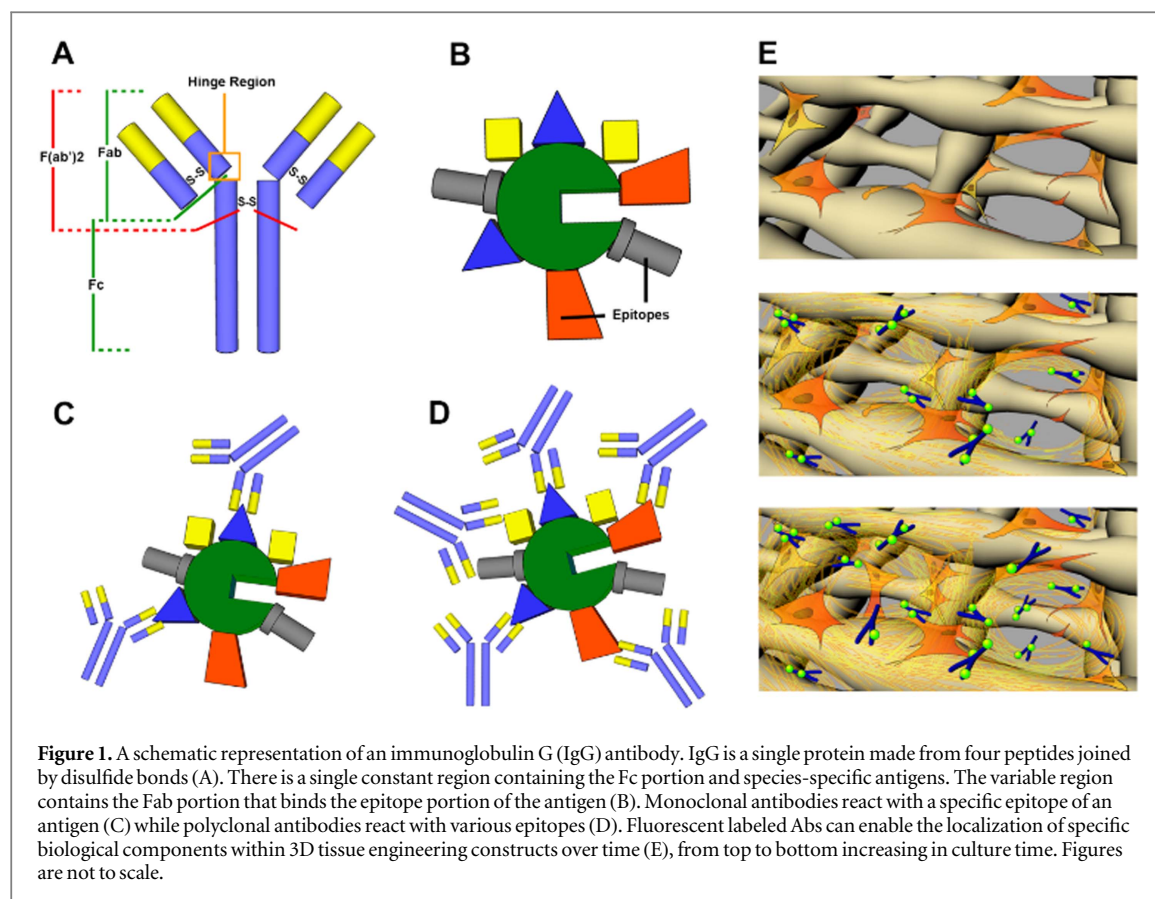
When tissue engineering strategies rely on the combination of three-dimensional (3D) polymeric or ceramic scaffolds with cells to culture implantable tissue constructs *in vitro*, it is desirable to monitor tissue growth and cell fate to be able to more rationally predict the quality and success of the construct upon implantation. Such a 3D construct is often referred to as a ‘black-box’ since the properties of the scaffolds material limit the applicability of most imaging modalities to assess important construct parameters. These parameters include the number of cells, the amount and type of tissue formed and the distribution of cells and tissue throughout the construct. Immunolabeling enables the spatial and temporal identification of multiple tissue types within one scaffold without the need to sacrifice the construct. In this report, we concisely review the applicability of antibodies (Abs) and their conjugation chemistries in tissue engineered constructs. With some preliminary experiments, we show an efficient conjugation strategy to couple extracellular matrix Abs to fluorophores. The conjugated probes proved to be effective in determining the presence of collagen type I and type II on electrospun and additive manufactured 3D scaffolds seeded with adult human bone marrow derived mesenchymal stromal cells. The conjugation chemistry applied in our proof of concept study is expected to be applicable in the coupling of any other fluorophore or particle to the Abs. This could ultimately lead to a library of probes to permit high-contrast imaging by several imaging modalities.

**1. Introduction**

Tissue engineering approaches for cartilage and bone regeneration often involve the use of three-dimensional (3D) synthetic polymeric or ceramic scaffolds with a certain mechanical integrity to mimic the natural native environment. Scaffolds can be implanted directly into the defect site, in which they are expected to attract cells and facilitate tissue ingrowth. Another approach is based on *in vitro* culture of scaffolds combined with autologous (stem) cells to be able to control culture parameters and therewith steer the cell fate and tissue growth into a desired lineage. In both cases, our understanding of the biological phenomena occurring within the pore network of 3D scaffolds is often limited. Real time monitoring of such phenomena offers a tool to also

advance the design of scaffolds which can better control cell fate and tissue growth. In particular, monitoring and being able to discriminate between different types of collagen would be important to further understand tissue morphogenesis in scaffolds. Collagen is the most abundant protein in mammals, representing about 30% of the total protein mass. There are currently 28 different collagen types discovered. Collagen type I is the most common and can be found in several tissues, while collagen type II is most found in articular cartilage [1]. However, current imaging techniques are hindered by the scaffolds limited transparency, scaffolds geometry and a lack of available methods to non-invasively assess cells and tissues functionality [2].

In the past few decades, the need for non-invasive monitoring methods has been widely acknowledged.



Several imaging methods have been opted to overcome transparency issues, such as ultrasound, nuclear, photoacoustic, x-ray and magnetic resonance (MR) based imaging modalities [3]. X-ray and MR imaging have shown to be promising tools in quantification of mineralization in bone tissue engineering [4, 5]. However, to date none of these methods enables the detection of multiple early tissue lineage markers with a single cell-resolution in 3D without the need to combine several imaging modalities.

The evolution of bio-conjugation chemistry has significantly contributed to the development of novel and successful methods to label multiple components within a single construct for imaging purposes. Antibodies (Abs) stand out as probes due to their specificity, affinity and versatility to target tissue specific components. Abs are proteins belonging to the immunoglobulin group and one of the most important defense mechanisms in vertebrate animals. In research, they are used as highly specific probes in order to identify and locate intra- and extracellular proteins, differentiate cell types and separate proteins [6]. Immunoglobulins are made from two identical heavy chain (*H*, 55–70 kDa, either  $\kappa$  or  $\lambda$ ) and two identical light chain (*L*, 24–25 kDa, either  $\gamma$ ,  $\epsilon$ ,  $\delta$ ,  $\alpha$ , or  $\mu$ ) peptides joined by disulfide bonds (figure 1(A)). The Fc portion of Abs is constant and specific to the animal in which the antibody is generated while the Fab portion is variable and binds to the epitope part of the antigen. A single antigen has multiple epitopes

(figure 1(B)) to allow Abs to recognize it. Monoclonal antibodies are homogeneous and bind to a single epitope on the antigen (figure 1(C)), whereas polyclonal antibodies are immunochemically dissimilar, having affinities to various epitopes of the same antigen (figure 1(D)). A possible application of Ab-based extracellular matrix (ECM) detection in 3D tissue engineered scaffolds is schematically presented in figure 1(E). Initially, only cells are present in the *in vitro* culture (upper figure). Over time, the cells will start to secrete specific ECM components such as collagen to which a targeted label will bind. With increasing concentration of ECM components, the binding of labeled Abs will also increase, resulting in a higher overall brightness of the substrate upon optical detection.

The direct conjugation of antibodies to detectable labels or agents allows the combination of the label properties with the high specificity and selectivity of antibodies [7]. Three main label types can be distinguished: Ab [8–11] or enzymes [12, 13] based detection with fluorophores, or particulate based labeling [6, 14]. Fluorescent immuno-labels enable the detection of specific components qualitatively and quantitatively by detecting light emitted from a fluorophore with a different wavelength than the light used to excite the fluorophore [6]. Antibody conjugated particles were developed already decades ago for the immunolabeling of specific ECM components in scanning electron microscopy analysis, although the required

sample processing made real-time monitoring impossible [14]. To date, particle based labels comprise a wide range of materials from heavy metals such as colloidal gold or small gold particles [14–16] to magnetic beads [17–19], radioactive components [9, 20, 21] and combinations of, for example, microbeads and covalently bound or entrapped fluorescent moieties [22].

The reactivity of an antibody is similar as for proteins determined by its amino acid composition and tertiary structure [23]. Understanding the functional groups available on an antibody is vital for finding the most appropriate conjugation method. Most Ab labeling strategies use one of the three types of available reactive groups. These groups comprise primary amines ( $-\text{NH}_2$ ), sulfhydryl groups ( $-\text{SH}$ ) and carbohydrates (sugars) [24, 25]. Primary amines are the most commonly used groups for conjugation, since they are abundant, widely distributed and easily modified due to their reactivity and location on the surface of the antibody [26]. The N-hydroxysuccinimidyl ester (NHS ester) reactive group is the most specific and efficient reagent to target primary amines [26]. The only drawback of this strategy is that it may cause a significant decrease in the antigen-binding activity of the antibody [27]. This is due to the presence of primary amines in that site, which can also be modified. Moreover, lysines, on which primary amines can be found, have a positive net charge while fluorophores have negative net charge, which may alter antibody conformation and activity upon conjugation. This decrease in activity is particularly noticeable when working with monoclonal antibodies or when attempting to achieve a higher degree of labeling (DOL) [23]. The second possible target labeling Abs is via sulfhydryls. Disulfide bonds are important in Ab function as they contribute to covalently connect the two Ab halves at the hinge region. Because disulfides in the hinge region are the most susceptible to reduction, a commonly utilized method to cleave these disulfides bridges is by introducing papain [23]. Papain induced cleavage splits the antibody into monovalent halves without changing the antigen-binding site and ensures consistent labeling at a defined location and degree. The third useful target for labeling nanoparticles with antibodies is carbohydrate moieties [23]. Labeling carbohydrates requires more steps than labeling amines because carbohydrates must first be oxidized to obtain reactive aldehydes. Recently, Shrestha *et al* performed an extensive study on the application of the three previously mentioned conjugation methods to conjugate a specific Ab to one type of fluorophore [23]. They concluded that amine based conjugation was by far the best technique due to its simplicity, rapidity, ease of operation, higher conjugate yield and lower costs. To enable the direct coupling of a label to the Ab via  $-\text{NH}_2$ ,  $-\text{SH}$  or  $-\text{CHO}$  groups to form a detectable Ab, functionalization of the label and/or Ab is required. Several well-known strategies to couple a label covalently to the reactive groups of the Ab are via

maleimide activation, active ester formation and biotin-avidin or biotin-streptavidin coupling [28, 29]. Furthermore, some non-covalently coupling methods are developed via hydrophobic interactions, electrostatic association with the particles coating, ligand coordination such as thiols and histidines and nickel mediated assembly of histidine to carboxyl groups.

The aim of this study was to develop a protocol for the conjugation of bone and cartilage specific Abs to be able to monitor ECM formation in 3D tissue engineered constructs. The probes have been developed considering that they should provide enhanced contrast in imaging modalities such as MRI, micro-CT or nuclear imaging for which transparency of the construct does not form an issue. Therefore, the conjugation chemistry should be compatible with particles that could be applied as contrast agents. Here, we chose to optimize and test anti-collagen-I and anti-collagen-II Abs conjugated with fluorescent probes as a proof of concept. We have validated the binding specificity with collagen type-I and collagen type-II on collagen type-I coated glass slides, native cartilage and bone tissue sections and on 3D tissue engineered constructs. We show that our conjugation protocol provides a sufficient DOL retaining an adequate antibody-antigen binding affinity to detect ECM components.

## 2. Materials and methods

### 2.1. Ab-dye conjugation procedure

Anti-collagen type-I (ab34710, Abcam<sup>®</sup>) and anti-collagen type-II (ab34712, Abcam<sup>®</sup>) (both polyclonal, produced in rabbit), the dyes ATTO 488 NHS-ester (ATTO-TEC GmbH) and ATTO-594 NHS-ester (ATTO-TEC GmbH) and Zeba<sup>™</sup> Micro Spin Desalting Columns (40 K MWCO, 75  $\mu\text{l}$ , Thermo Fisher Scientific) were used. Prior to use, the buffer in the columns was exchanged to phosphate buffered saline (PBS) with 0.02 M sodium azide according to manufactures instruction. Sodium azide was used as a preservative to inhibit the growth of contaminants such as bacteria or fungi in the antibody solutions. Before the conjugation step, 10  $\mu\text{l}$  of antibody solution was passed through the column once in order to exchange the antibody buffer solution to PBS solution. The product was collected in an amber colored 1.5 ml eppendorf tube. To adjust the pH to the optimal conjugation value (pH = 8.3), sodium bicarbonate (1 M, pH = 8.3) was added to the antibody solution. On the first use, the dye was dissolved to 2  $\text{mg ml}^{-1}$  in anhydrous DMSO according to manufactures instruction, aliquot and stored at  $-20^\circ\text{C}$  for further experiments. In order to obtain the desired DOL between 2 and 3 labels per antibody (higher DOL results in non-functional antibodies), approximately 0.1  $\mu\text{l}$  of dye solution were added to the antibody solution, mixed and incubated for 1 h at RT with continuous shaking



on an oscillating  $x$ - $y$  plate. The conjugate solution was then passed three times through new desalting columns to remove any unconjugated dye. An ND100 spectrophotometer (Nanodrop Technologies, USA) was utilized to analyze the obtained concentration and ensure that the reaction occurred as expected. If required, BSA was added to increase the solution concentration above  $1 \text{ mg ml}^{-1}$ . The anti-collagen type-I conjugated ATTO 488 dye will further be referred to as 'anti-col-I'. The anti-collagen type-II conjugated ATTO 594 dye will further be referred to as 'anti-col-II'. The conjugate solution was stored at  $4^\circ\text{C}$ . As positive control to assess conjugates functionality, the same anti-collagen type-I or type-II antibody and a secondary anti-rabbit antibody conjugated to an Alexa-Fluor 488 fluorophore were used in several experiments as a positive control and will be further referred to as 'control probe'.

## 2.2. Conjugate characterization

The efficiency of the procedure and DOL of the antibody-dye conjugate were determined with a ND1000 spectrophotometer (Nanodrop Technologies, USA). The following equation (equation (1)), based on the Lambert–Beer law, was applied to calculate the DOL:

$$\begin{aligned} \text{DOL} &= \frac{[\text{dye}]}{[\text{protein}]} = \frac{A_{\text{max}}/\varepsilon_{\text{max}}}{A_{\text{prot}}/\varepsilon_{\text{prot}}} \\ &= \frac{A_{\text{max}}/\varepsilon_{\text{prot}}}{(A_{280} - A_{\text{max}} * \text{CF}_{280}) * \varepsilon_{\text{max}}}, \end{aligned} \quad (1)$$

where:  $A_{280}$  is the absorbance at 280 nm,  $A_{\text{max}}$  is the absorbance at  $\lambda_{\text{max}}$ ,  $\lambda_{\text{max}}$  is the wavelength where the absorbance reaches its maximal value, CF is the correction factor which depends on the dye's spectral properties,  $\varepsilon_{\text{prot}}$  is the molar extinction coefficient ( $\text{M}^{-1} \text{ cm}^{-1}$ ) of the pure protein at 280 nm and  $\varepsilon_{\text{max}}$  is the molar extinction coefficient ( $\text{M}^{-1} \text{ cm}^{-1}$ ) of the dye at its absorbance maximum. According to literature, an ideal DOL sets between 2 and 8 depending on the materials and methods utilized. Thus, we aimed at a practical DOL of 4. SDS-PAGE was applied to get an impression of the purity of the conjugate. The gel was run at 70 V for stacking and then shifted to 30 mA until separation was completed.

## 2.3. Scaffold fabrication

Scaffolds were fabricated as described before, from 300PEOT55PBT45 (300/55/45) (PolyVation, The Netherlands), a block copolymer composed of poly(ethylene oxide terephthalate) (PEOT) and poly(butylene terephthalate) (PBT) with a weight ratio of 55–45 for the two components, respectively, and a molecular weight (MW) of the starting poly(ethylene glycol) (PEG) segments of 300 Da used in the co-polymerization process [30].

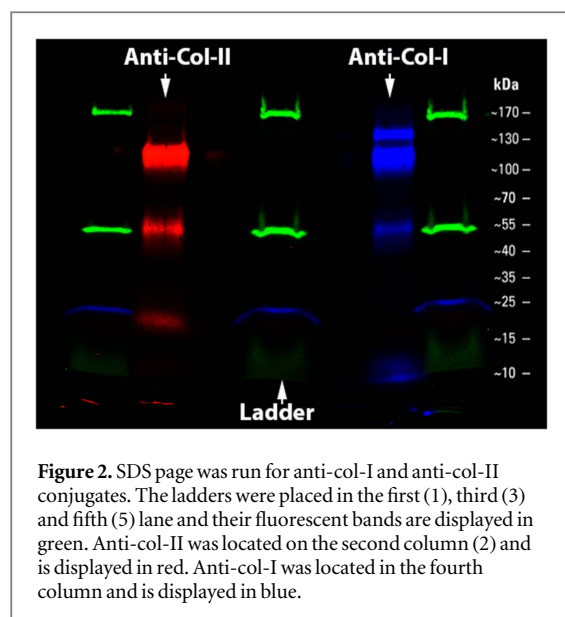
For electrospun (ES) scaffolds, a 28% (w/v) solution of 300/55/45 was prepared in a mixture of chloroform

( $\text{CHCl}_3$ )-1,1,1,3,3,3-hexafluoro-2-propanol (HFIP) with a volume ratio of 78:22. The solution was left stirring at room temperature overnight. Subsequently, the solution was loaded into a syringe and pumped at a controlled flow rate using a syringe pump (KDS 100, KD Scientific). PTFE tubing was used to connect the syringe and a needle to which a high voltage was applied (Gamma High Voltage Research Inc., FL, USA). A stainless steel collector, covered by aluminum foil, was used to collect the ES scaffold. A voltage of 12 kV was applied to a 1.2 mm needle, which was placed at 25 cm from the collector. The flow rate was set at  $15 \text{ ml h}^{-1}$ , temperature to  $25^\circ\text{C}$ , and humidity to 30%. Upon reaching a sheet thickness of approximately  $200 \mu\text{m}$  the electrospinning process was stopped and disks with a diameter of 15 mm were punched out from the sheet.

Additive manufacturing by means of fused deposition modeling with a bioscaffold (SysENG, Germany) was used to fabricate 3D plotted scaffolds, as described before [31, 32]. The cylindrical shaped scaffolds had diameter of 8 mm a height of 3 mm, a fiber-to-fiber distance of  $1000 \mu\text{m}$ , a fiber diameter of approximately  $200 \mu\text{m}$ , and a layer thickness of  $150 \mu\text{m}$ .

## 2.4. Cell culture

Bone marrow aspirates were obtained from donors after written informed consent and human bone marrow stromal cells (hMSCs) were isolated and proliferated as previously described [33]. In short, 20 gauge needles were used to re-suspend the aspirates, which were then plated at a density of  $5 \times 10^5 \text{ cells cm}^{-2}$ . Cells were cultured in hMSC proliferation medium, for which the composition is described in the supplementary data is available online at [stacks.iop.org/BMM/13/044102/mmedia](https://stacks.iop.org/BMM/13/044102/mmedia), at  $37^\circ\text{C}$  in a humid atmosphere with 5%  $\text{CO}_2$ . The culture medium was refreshed three times a week. After monolayer expansion, cells were either cryopreserved or used for further subculturing. Passage 3 hMSCs were detached from the culture flasks by trypsinization upon reaching 70% confluence. Basic medium contained  $\alpha$ -minimal essential medium (Life Technologies), 10% heat-activated fetal bovine serum (Cambrex), 0.2 mM ascorbic acid (Asap; Life Technologies), 2 mM L-glutamine (Life Technologies), 100 units  $\text{ml}^{-1}$  penicillin (Life Technologies), 10  $\mu\text{g ml}^{-1}$  streptomycin (Life Technologies). Proliferation medium consisted of basic medium supplemented with  $1 \text{ ng ml}^{-1}$  basic fibroblast growth factor (Neuromics, Acris Antibodies, Germany). Osteogenic medium consisted of basic medium supplemented with 10 nM dexamethasone. Chondrogenic medium contains Dulbecco's modified essential medium (D-MEM) (Life Technologies) with 0.2 mM ascorbic acid, 100 units  $\text{ml}^{-1}$  penicillin (Life Technologies), 10  $\mu\text{g ml}^{-1}$  streptomycin (Life Technologies), 50  $\mu\text{g ml}^{-1}$  ITS-premix, 100  $\mu\text{g ml}^{-1}$  sodium pyruvate, 10  $\text{ng ml}^{-1}$  TGF- $\beta$ 3 and 1  $\mu\text{M}$  dexamethasone. TGF- $\beta$ 3 and dexamethasone



were freshly added every time that the media was changed during culture.

All scaffolds were sterilized by immersion in 70% ethanol for at least 2 h, washed multiple times with sterile PBS and subsequently placed into a non-treated 24 well plate (NUNC). The constructs were then incubated at 37 °C in a humid atmosphere with 5% CO<sub>2</sub> overnight in basic medium to pre-wet them. To prevent the ES scaffold discs from floating, rubber O-rings (Eriks BV, The Netherlands) were used to hold the discs in place. After incubation, the medium on the scaffold was aspirated, and 75 000 cells and 250 000 cells were seeded onto each ES scaffold and plotted scaffolds, respectively. After 7 d of culture in proliferation medium, the media was replaced with chondrogenic and osteogenic medium to start differentiation and cultured for additional 14 d.

## 2.5. Immunostaining

Glass slides with a diameter of 18 mm were immersed in 10% acetic acid in ethanol for 30 min, followed by 100% ethanol rinses and dried in the oven at 50 °C. Collagen type I from rat tail (BD Biosciences, Two Oak Park, Bedford, USA) was diluted from a stock concentration of 11.59 to 2 mg ml<sup>-1</sup> in a 0.02 M acetic acid solution. Glass slides were placed in a non-treated 12 well plate (NUNC) and 250 µl of collagen solution was added. After 1 h incubation at 37 °C, glass slides were washed three times with PBS to remove any remaining acetic acid. Prior to Ab incubation, slides were blocked with a buffer (0.1% Triton X-100, 1% BSA in PBS). Afterwards, they were incubated with the fluorescently conjugated primary Abs, at 4 °C, overnight. After incubation, slides were washed three times with PBST (0.1% Tween-20 in PBS) for 5 min each. The presence of collagen coating was further confirmed by picosirius red staining (supplementary figure 1(C)).

A 6 weeks old post-mortem rat was kindly provided by the department of Biomedical Signals and

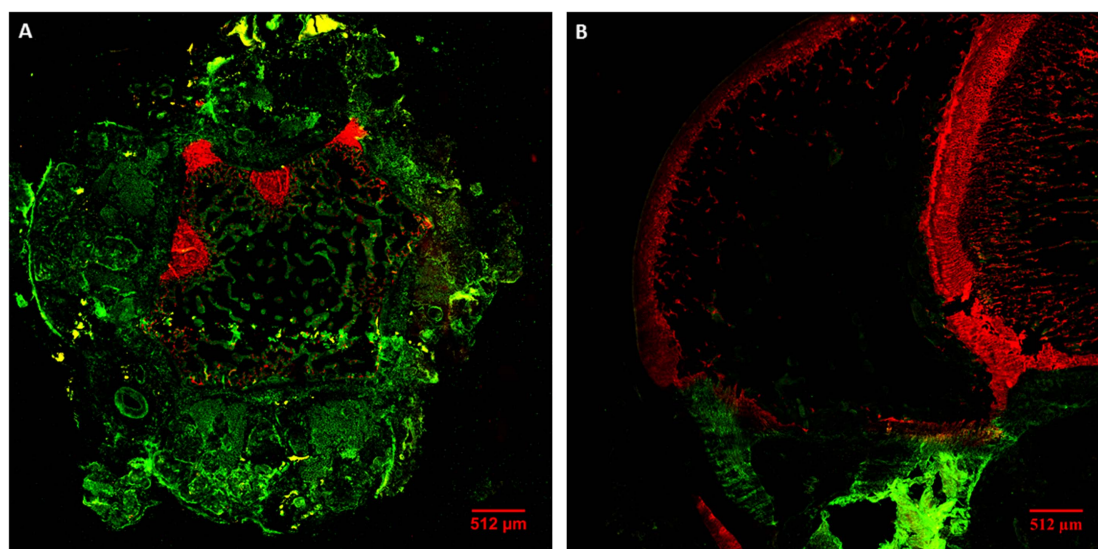
Systems (BSS, MIRA institute, University of Twente). Animal use was regulated by national laws on animal experiments after approval from a local ethical committee. The tail and bone of the rat was isolated and immersed in 12.5% EDTA, pH = 8, for 5 weeks to decalcify the tissues. EDTA was changed three times a week. A series of increasing ethanol concentrations (60%, 70%, 80%, 90%, 96%, 100% × 2), 2 d per concentration, was used to dehydrate the tissue. Subsequently, the tissue was immersed in butanol overnight and in eosin for 1 h to stain them with a pink color to be able to localize the sample after embedding. Specimens were embedded in paraffin for 5 d in a holder at 60 °C. Paraffin was changed after 2 d to remove any traces of butanol. The sections were cut at room temperature with 5 µm intervals. Prior to Ab incubation, specimens were blocked with a buffer (0.1% Triton X-100, 1% BSA in PBS). Incubation was performed with the fluorescently conjugated primary Abs, at 4 °C, overnight. After incubation, specimens were washed three times with PBST for 5 min each. Afterwards, specimens were incubated in a solution of 0.1% Sudan Black B in 70% Ethanol for 20 min to mask bone autofluorescence and washed three times with PBS for 5 min each.

After 14 d of culture in differentiation media, the cultured samples were washed once with PBS, fixed with paraformaldehyde (4% in PBS) for 15 min and stored in PBS at 4 °C. Prior to Ab incubation, constructs were blocked with a buffer (0.1% Triton X-100, 1% BSA in PBS). Samples were incubated with both anti-col-I and anti-col-II Abs, at 4 °C overnight. Control samples were incubated with primary Abs (ab34710 and ab34712, Abcam®), at room temperature, for 1 h. Then, constructs were washed once with PBS and incubated with the corresponding secondary antibody (anti-rabbit antibody conjugated to an Alexa-Fluor 488 fluorophore) overnight, at 4 °C. After incubation, constructs were washed three times with PBST (0.1% Tween-20 in PBS) for 5 min each. Afterwards, samples were incubated in a solution of 0.1% Sudan Black B in 70% Ethanol for 20 min to remove polymer autofluorescence and then washed three times with PBS for 5 min each. Cell nuclei labeling was performed with 4',6'-Diamidin-2'-phenylindoldihydrochlorid (DAPI, 100 ng ml<sup>-1</sup> in PBS, Sigma) and actin filaments were stained with Phalloidin to visualize the cytoskeleton. Samples were imaged using a Nikon A1 confocal microscope (Nikon Instruments Inc.).

## 3. Results and discussion

### 3.1. Ab and dye concentration by spectrophotometry

To quantify the DOL after conjugation of the fluorescent probe to the Ab, the absorbance spectrum was analyzed with a ND1000 spectrophotometer



**Figure 3.** Fluorescent microscopy images of (A) tail and (B) bone sections, stained by anti-col-I (green) and anti-col-II (red). Scale bar: 512  $\mu\text{m}$ .

(Nanodrop Technologies, USA). The CFs for ATTO 488 and for ATTO 594 were provided by the supplier and found to be 0.1 and 0.51, respectively. The  $\epsilon_{\text{max}}$  for ATTO 488 and for ATTO 594 were also provided by the supplier and were 90 000 and 120 000  $\text{M}^{-1} \text{cm}^{-1}$ , respectively. The molar extinction coefficient for the AB (IgG) was set at 210 000  $\text{M}^{-1} \text{cm}^{-1}$ . The path length in all measurements was set at 1 cm. For the anti-col-I the  $A_{280}$  was found to be 0.877, while  $A_{\text{max}}$  was found to be 1.953 at approximately 500 nm. With these values the DOL was determined according to equation (1) and found to be around 6. For, anti-col-II conjugated to ATTO 594 the  $A_{280}$  was found to be 2.115, while  $A_{\text{max}}$  was found to be 0.435 at approximately 500 nm. With these values the DOL was determined according to equation (1) and found to be around 4. Both values are well under the optimal range for DOL which sets between 2 and 8. An excessively high DOL may denature the antibodies or reduce its antigen-binding activity, decreasing its functionality. A low DOL will result in a low signal. Further analysis of the probe functionality will be accessed in the following section.

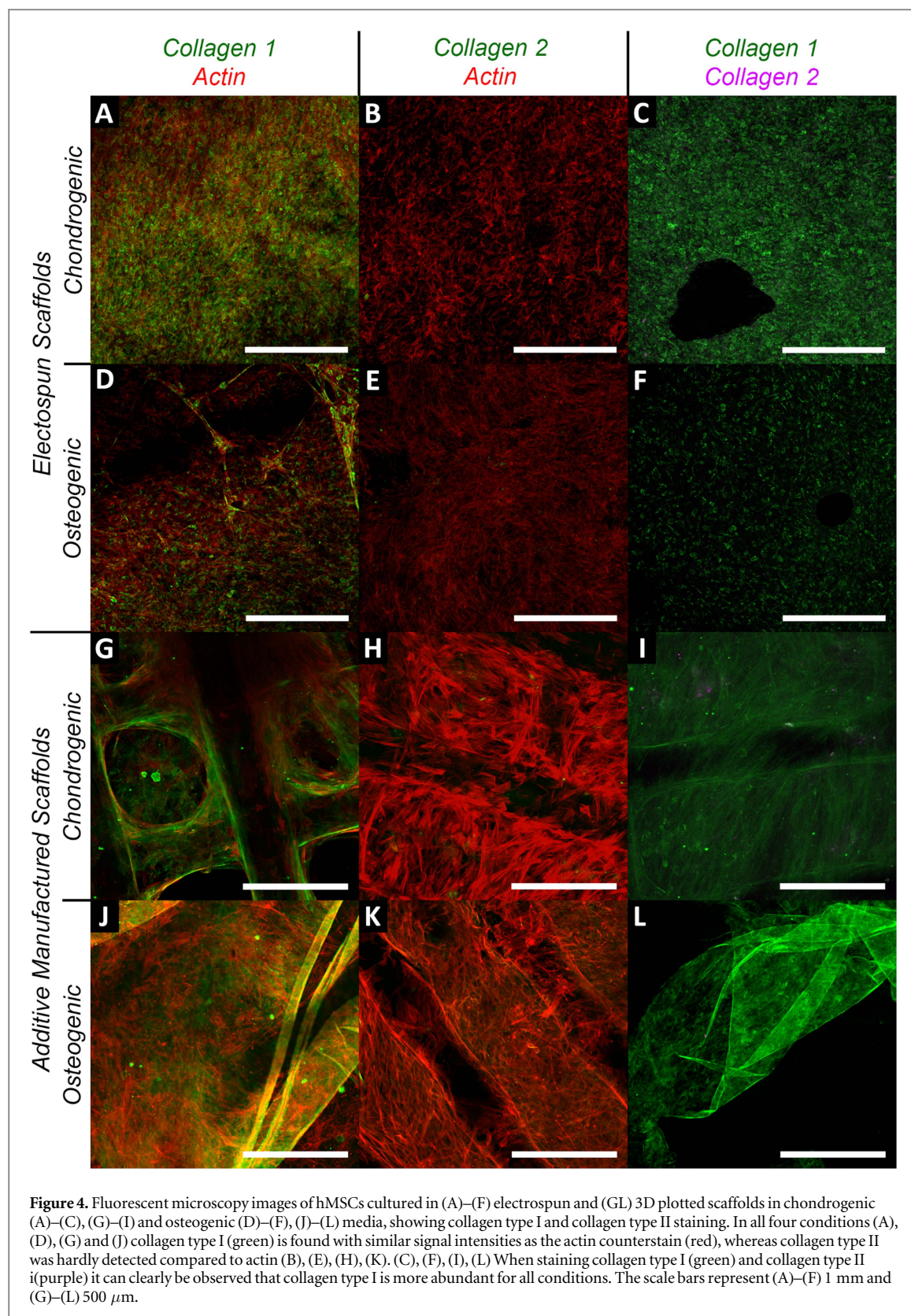
SDS-PAGE was performed before adding BSA to the conjugate solution to get insight in the purity of the probes solution (figure 2). In both antibody conjugated dyes, a strong fluorescent band was observed in the top of the gel, where higher MW molecules were retained. Two other bands were observed at lower MW. However, these bands seemed significantly weaker compared to the ones in the top layer of the gel. The probe solution was therefore considered sufficiently purified with just low amounts of unconjugated label and Abs.

### 3.2. Cell culture

The ultimate goal of the probe developed here was to be applied on culture systems to monitor and characterize tissue formation in 3D scaffolds *in vitro*. Firstly, the antigen affinity of the anti-col-I was assessed on a pure collagen type-I coating (Rat) and compared to the affinity of the commercially available control probe (supplementary figure 1). It was observed that both probes showed high binding affinity with the collagen coating whereas no label was detected on the surface of the glass slide. This was done to ensure that the functionality of both conjugated probes was preserved. The different net charges of the fluorophore (−1) and the amine groups (+1) and the possibility of fluorophore conjugation to the Ab antigen-site could have reduced its final functionality, even though the DOL was well between optimal values. Anti-col-I and anti-col-II were further tested on a histological section of demineralized rat tail and bone tissue to get insights in non-specific binding (figure 3), further confirming the successful synthesis of the molecular probes. All structures were blocked with a BSA based blocking buffer to prevent non-specific binding and secondary Abs were applied to probe successful blockage (supplementary figure 3). Moreover, both control probes and anti-col-I and anti-col-II showed similar detection sites (supplementary figure 3).

The two probes were, then, applied on hMSCs cultured for 14 d in osteogenic or chondrogenic medium on ES and 3D plotted scaffolds (figure 4). ES scaffolds showed randomly oriented fibers with a non-uniformly distributed average diameter of  $3.06 \pm 1.95 \mu\text{m}$ , with a minimum diameter of approximately  $1 \mu\text{m}$  and maximum fiber diameter of  $8 \mu\text{m}$ . Conversely, 3D plotted





scaffolds showed well oriented fibers with an average fiber diameter of  $214.5 \pm 27.1 \mu\text{m}$  and a fiber spacing of  $498.4 \pm 46 \mu\text{m}$ .

Results showed that collagen type I was abundantly produced by hMSCs in both scaffold types and in both media. Collagen type II was hardly expressed in osteogenic medium and minimally produced in

chondrogenic medium in both scaffold types, thus suggesting better differentiation in osteogenic conditions than in chondrogenic conditions. In our previous studies, we have also found similar results at a genetic level, where it was clear that hMSCs on PEOT/PBT scaffolds showed upregulated early markers for both osteogenesis and chondrogenesis, accompanied



by higher expression of alkaline phosphatase activity and glycosaminoglycans, respectively [30].

The anti-co-II probe was also tested against a control probe in osteogenic conditions on 3D plotted scaffolds (supplementary figure 2). The circular pattern formed by the seeded hMSCs has shown to be typical for tissue formation in this type of scaffolds, where cells initially attach on the struts of the scaffolds and over time close the pore. This pattern of tissue growth is one of the processes that can be considered important to assess during culture to monitor the type and morphology of the ECM that was produced. Collagen type-I was found both with anti-col-I and with the control probe. In both samples, the collagen type-I showed to be distributed over the areas where cells were located as well. Therefore, the newly developed probes of this study can be considered successfully applied. The high DOL obtained in this study could have interfered with the binding affinity of the Ab to the antigen, which could explain the lower brightness of the anti-col-I compared to the control probe. The stability of the label was not expected to result in this lower brightness, since the ATTO-dyes applied in anti-col-I and anti-col-II are generally more stable than the fluorophore coupled the control probe. Yet, there are some other advantages for the conjugated antibodies. They do not require a secondary incubation with a secondary antibody labeled antibodies, which reduces the amount of protocol steps and unspecific secondary antibody binding. These conjugated-antibodies allow the use of two or more antibodies raised in the same animal, e.g. useful in this study due to the limited availability of collagen II antibodies. Finally, they allow the conjugation of any fluorophore or nanoparticle with a NHS-ester, especially important in cases in which they are not commercial available and readily conjugated for secondary incubation.

In theory, the conjugation chemistry applied on this Ab allows for the coupling to any other kind of labeling particle, thus being applicable also to the near infrared spectrum. Applying other imaging modalities would permit imaging of collagens at greater depths without the need to stain the cells. By coupling collagen type detection to a responsive scaffold material, for example a sort of actuating system to locally administer certain stimuli, more insight in collagen production and collagen orientation could be realized.

## 4. Conclusion

The main objective of this study was the optimization and validation of a method that could be used to functionalize particles and fluorophores to be applied in several imaging modalities. Based on a concise review on Abs and available conjugation strategies, the most promising method for application in 3D tissue engineered scaffolds was chosen and optimized. Both

anti-collagen type-I and anti-collagen type-II Abs were successfully conjugated to their respective dyes. The anti-col-I probe showed a similar selectivity to a control probe targeting collagen type-I in all tests performed, whereas anti-col-II revealed no maturation of chondrogenic differentiation of hMSCs in both ES and 3D plotted scaffolds. Yet, from the successful collagen type-I and type II detection, we conclude that the conjugation via amines offers an easy and feasible chemistry to obtain specific anti-collagen-I and -II Abs coupled to several types of fluorophores and particles.

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